

The background of the slide features a large, faint watermark of the Rutgers University seal. The seal is circular and contains the text 'RUTGERS THE STATE UNIVERSITY OF NEW JERSEY' around its perimeter. The central part of the seal is a sunburst design.

# RUTGERS

Robert Wood Johnson  
Medical School

## Biomarkers of MS therapies

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Professor and Chairman  
Department of Neurology

# Disclosures

- *Consultant:* Bayer, Biogen, EMD Serono, Teva, Novartis, Acorda
- *Investigator/Research Grants:* TEVA, Biogen

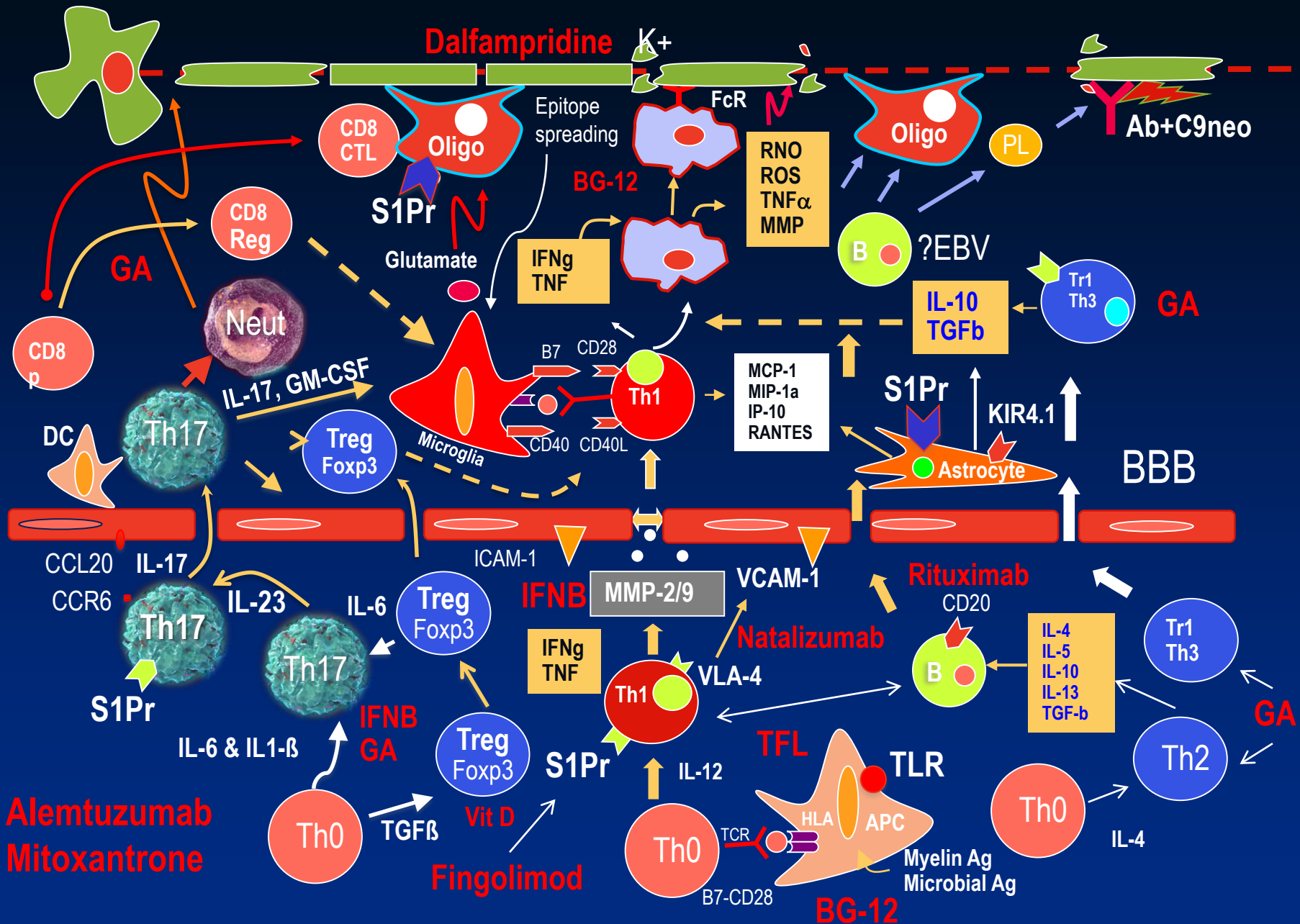
# Significance of biomarkers discovery in MS

- Early diagnosis
- Predict clinical course (Benign, RR, PP)
- Assessment of disease activity
- Pathological phenotype
- Surrogate outcome in clinical trials
- Prediction of treatment response

# **MS: Disease heterogeneity is a challenge to biomarkers discovery**

- Variable clinical course
- Pathological heterogeneity
- Inconsistent immune findings
- Variable response to DMDs

# Immunopathogenesis of MS



# BIOMARKERS OF TREATMENT RESPONSE

- Understand drug MOA
- Target molecules involved in the drug's MOA as potential biomarkers
- Genomics
- Proteomics

# Interferons Biomarkers

# Potential Mechanisms of Action of IFN- $\beta$ in MS

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- Anti-proliferative effect
- Blocking T-cell activation
- Apoptosis of autoreactive T-cells
- IFN-g antagonism
- Cytokine shifts
- Blood-brain barrier
- Antiviral effect





Contents lists available at [SciVerse ScienceDirect](#)

## Journal of Neuroimmunology

journal homepage: [www.elsevier.com/locate/jneuroim](http://www.elsevier.com/locate/jneuroim)



### Immune response during interferon beta-1b treatment in patients with multiple sclerosis who experienced relapses and those who were relapse-free in the START study

Suhayl Dhib-Jalbut <sup>a,\*</sup>, Sumandeep Sumandeep <sup>a</sup>, Reuben Valenzuela <sup>a</sup>, Kouichi Ito <sup>a</sup>,  
Payal Patel <sup>b</sup>, Mark Rametta <sup>b</sup>

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# Overview of Exploratory Outcomes

## Exploratory Outcomes (Immune Parameters)

### Blood Brain Barrier Markers

- sVCAM
- TIMP-1
- MMP-2
- TIMP-1/MMP-2 Ratio

### Cytokines

<b>Pro-Inflammatory Cytokines</b>	•IFN-gamma	•TNF-alpha	•IL-17
<b>Anti-Inflammatory/Regulatory Cytokines</b>	•TGF-beta •IL-4	•IL-10 •IL-13	•IL-27
<b>Ratios</b>	•IL-4/IFNg •IL-13/IFNg	•IL-10/IFN-g •IL-10/IL-17 ratio	

### Neurotrophic factor expression

- Brain-derived neurotrophic factor (BDNF)
- Nerve growth factor (NGF)

### Chemokines (Expression)

- CXCR3 (Th1)
- CCR4 (Th2)

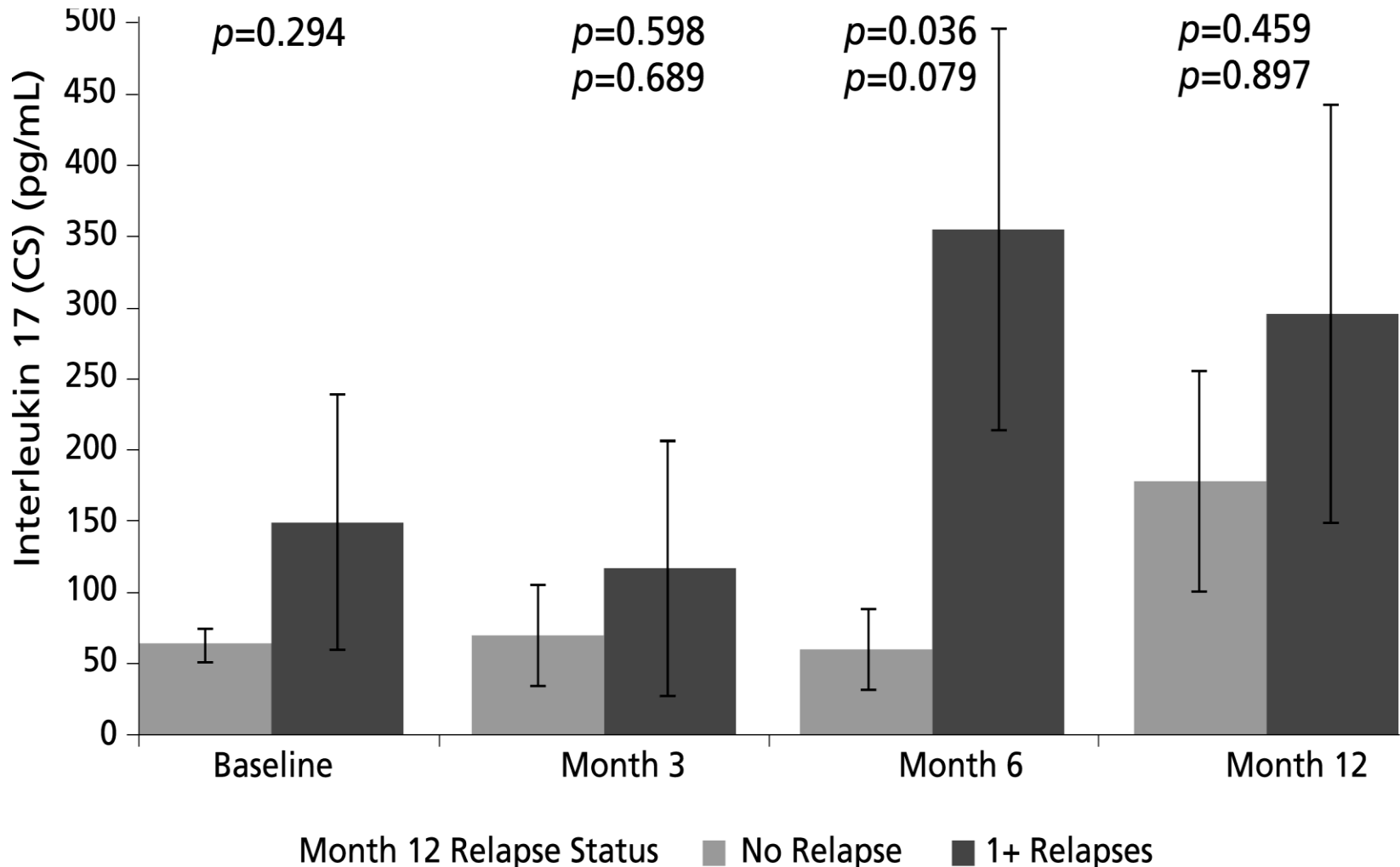
# Interferon therapy: **Baseline** molecules in relapsing and relapse-free subjects

Immune Parameter	Time point	Statistical Parameter	Relapse Free Subset	≥1 Relapses Subset	<i>p</i> -value
IL-10 (CS)	Baseline	N	14	11	0.119
		mean	26.739	70.276	
		SD	21.112	98.268	
		median	20.360	47.450	
IL-10/IFN-gamma ratio (CS)	Baseline	N	14	11	0.074
		mean	0.067	0.380	
		SD	0.057	0.626	
		median	0.059	0.160	
IL-4/IFN-gamma ratio (CS)	Baseline	N	14	11	0.025
		mean	0.024	0.085	
		SD	0.021	0.091	
		median	0.018	0.056	
BDNF serum	Baseline	N	14	11	0.011
		mean	675.6	525.0	
		SD	137.86	132.11	
		median	666.3	554.5	

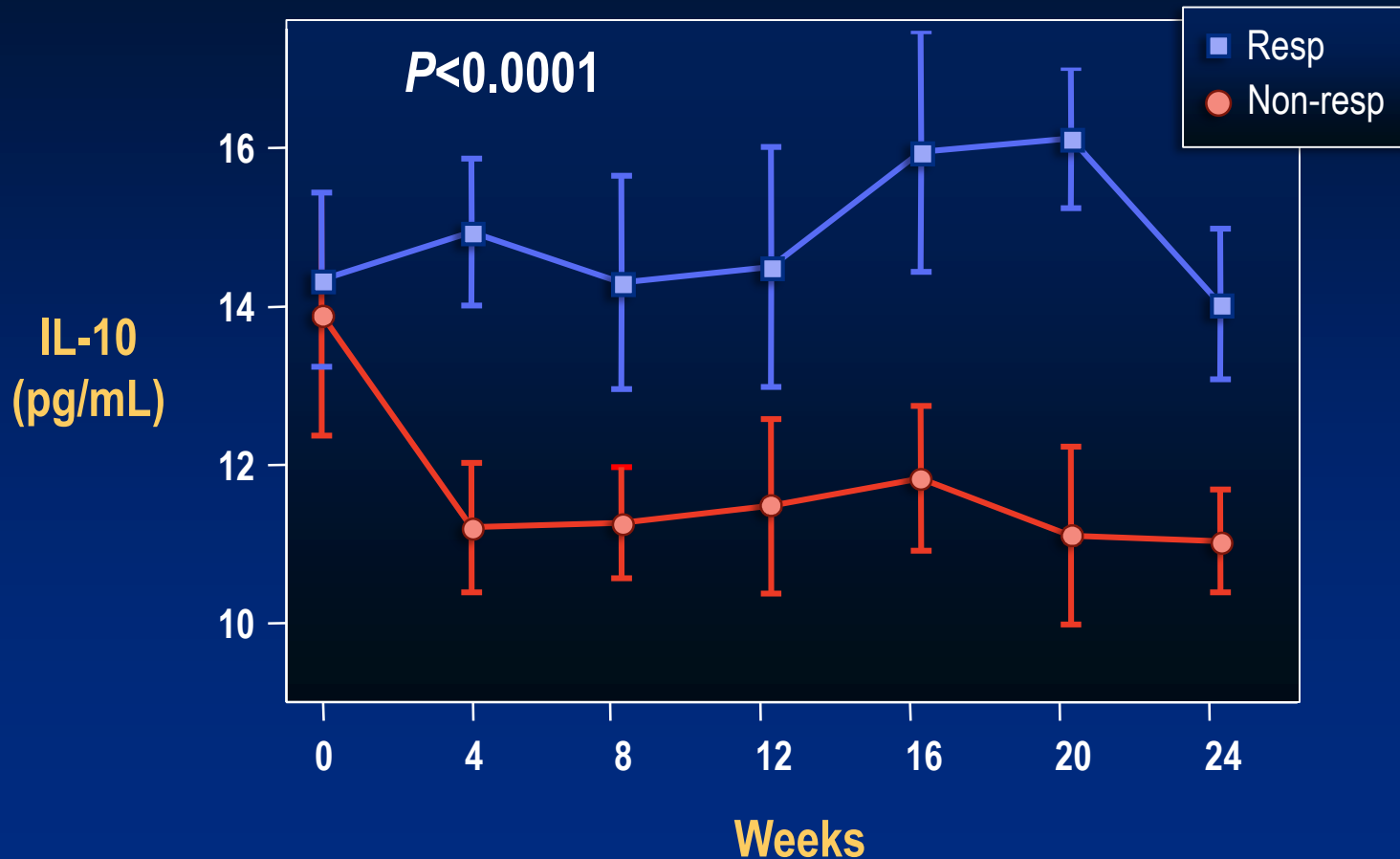
## Immune markers during IFN $\beta$ -1b treatment in relapsing and relapse-free subjects

Immune parameter	Time point (months)	Relapse-free subset Mean (SD)	$\geq 1$ Relapse subset Mean (SD)	<i>p</i> -value
IL-17 (CS)	6	59.78 (100.91)	355.21 (466.87)	0.036
CXCR3 (PBMC)	3	24.9 (6.92) <sub>r</sub>	33.2 (12.25)	0.042
IL-4 (CS)	3	13.43 (7.05)	20.88 (31.60)	0.046
BDNF serum	3	768.9 (142.7)	631.8 (148.0)	0.028

# IFN-B: Mean IL-17A Levels by Month 12 Relapse Status



# Serum IL-10 Levels During 24 Weeks Of IFN-beta Therapy



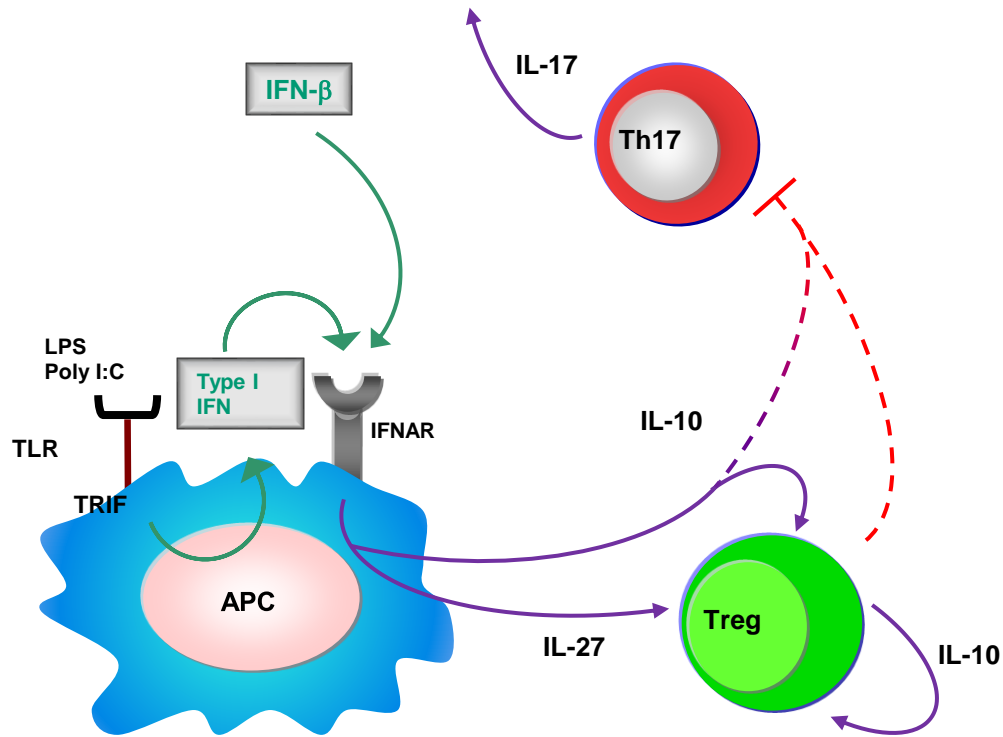
# Association Between Expression of Multiple Biomarkers and IFN Treatment Response

- Single biomarker expression is not a good predictor of therapeutic response
- Combinations of 2-3 biomarkers better predict therapeutic response
  - Gene combinations with high predictive value:
    - Caspase-2/IL-10/IRF-2
    - Caspase-2/IL-10/JNK-1
    - Caspase-2/IL-10
    - Caspase-2/IL-10/IL-12R $\beta$ 1

Caspase-2 involved in cell death; IL-10 involved in proliferation of T and B cells; IRF-2 involved in activation of IFN $\alpha$  and  $\beta$ ; JNK-1 involved in T cell differentiation and cell death; IL-12 R $\beta$ 1 binds IL-12

IFN = interferon

# Signaling via IFN- $\beta$ regulates Th17 differentiation



Prod'homme, T. and Zamvil, S.S. *Nat. Med.* 14: 614-616 (2008)

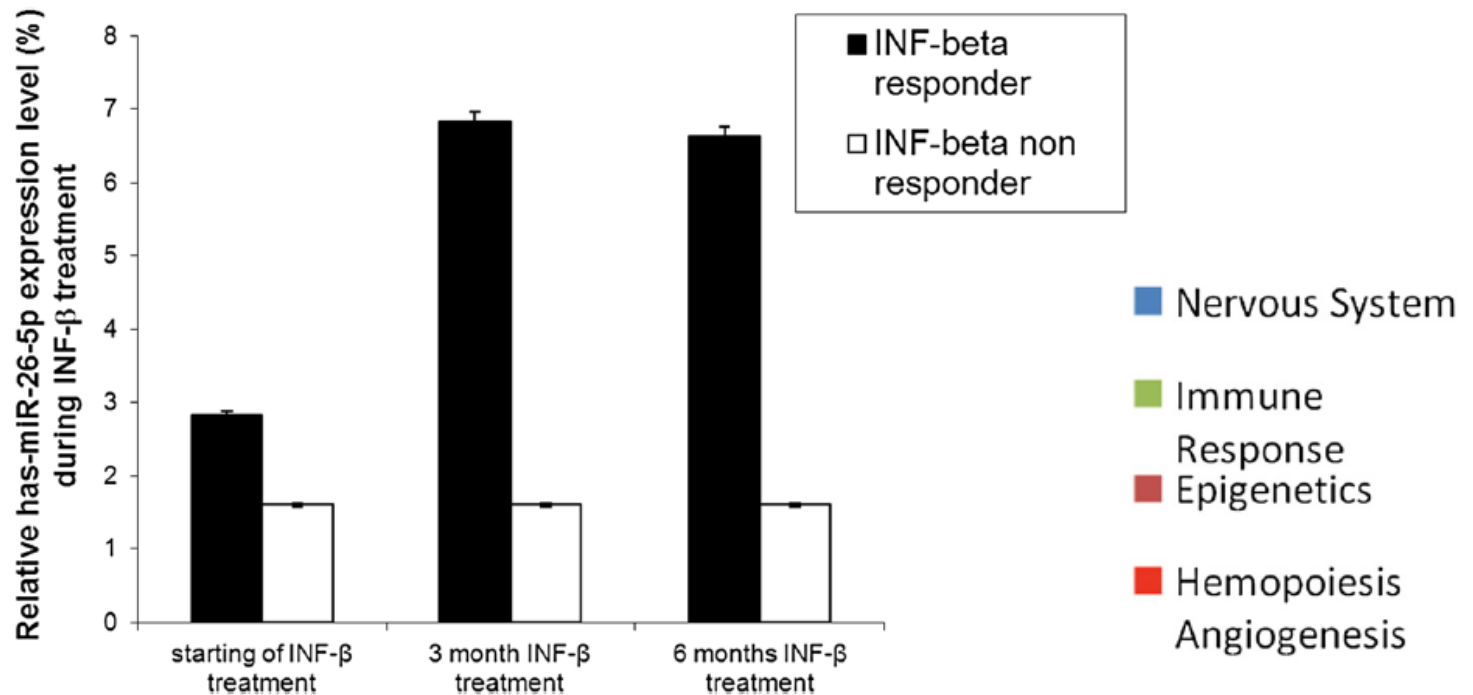


# IFN- $\beta$ Molecular Signatures Studies

- Weinstock-Guttman J. *Neuroimmunol.* 2008 Dec 15;205(1-2):113-25. Epub 2008 Oct 23
- Comabella et al. *Brain.* 2009 Dec;132(Pt 12):3353-65.
- Rudick et al. *PloS one.* 2011;6:e19262
- Croze et al. *Pharmacogenomics J.* 2012

# Small non-coding RNA signature in multiple sclerosis patients after treatment with interferon- $\beta$

Bruna De Felice<sup>1,4\*</sup>, Paolo Mondola<sup>2</sup>, Anna Sasso<sup>2</sup>, Giuseppe Orefice<sup>2</sup>, Vincenzo Bresciamorra<sup>2</sup>, Giovanni Vacca<sup>2</sup>, Elio Biffali<sup>3</sup>, Marco Borra<sup>3</sup> and Raimondo Pannone<sup>3</sup>

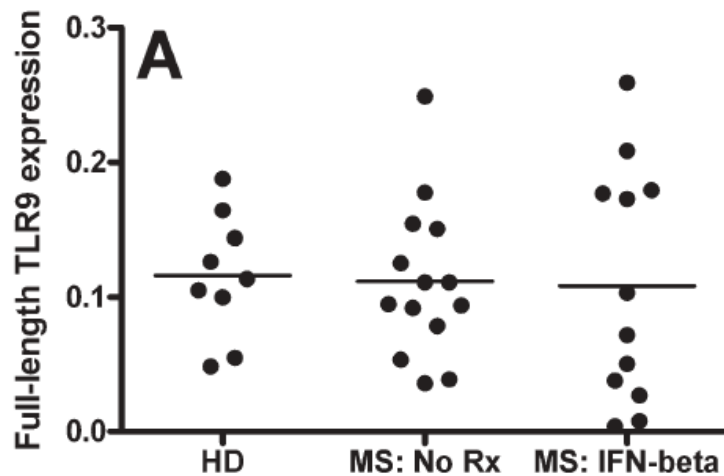


**Figure 2 MiR-26a-5p expression levels in INF- $\beta$  treated MS patient leukocytes.** The expression of mir-26a-5p was studied in the peripheral blood from 40 IFN- $\beta$  treated responder RRMS patients and from 10 IFN- $\beta$  treated non-responder patients, at starting, at 3 and 6 months IFN- $\beta$  treatment by microRNA assay-based quantitative RT-PCR following the  $2^{-\Delta\Delta CT}$  method. RNU6B was utilized for an endogenous reference to standardize microRNA expression levels. The results were expressed as relative expression levels after calibration with the universal reference data.  $P < 0.05$ .

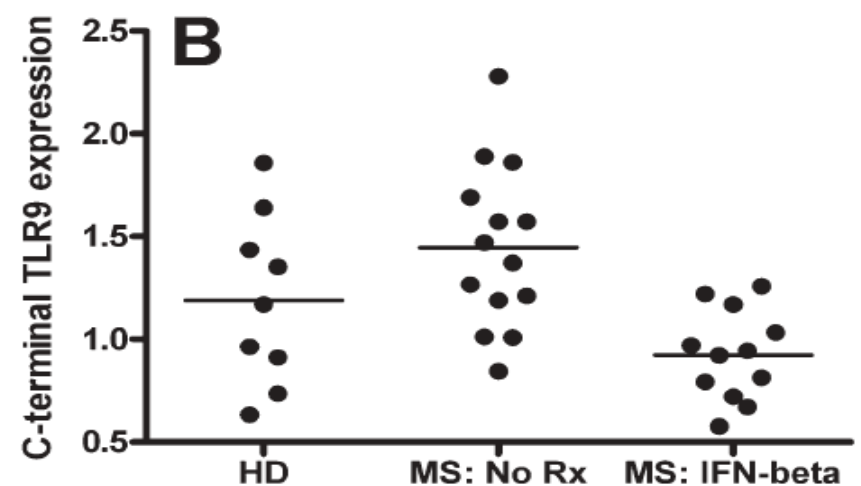
# Interferon-beta Inhibits Toll-Like Receptor 9 Processing in Multiple Sclerosis

Konstantin E. Balashov, MD, PhD,<sup>1,2</sup> Latt Latt Aung, MD,<sup>1</sup>  
Adi Vaknin-Dembinsky, MD, PhD,<sup>2</sup> Suhayl Dhib-Jalbut, MD,<sup>1</sup>  
and Howard L. Weiner, MD<sup>2</sup>

IFN-beta does not affect full-length TLR9 protein



IFN-beta inhibits TLR9 processing





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# Journal of Neuroimmunology

journal homepage: [www.elsevier.com/locate/jneuroim](http://www.elsevier.com/locate/jneuroim)

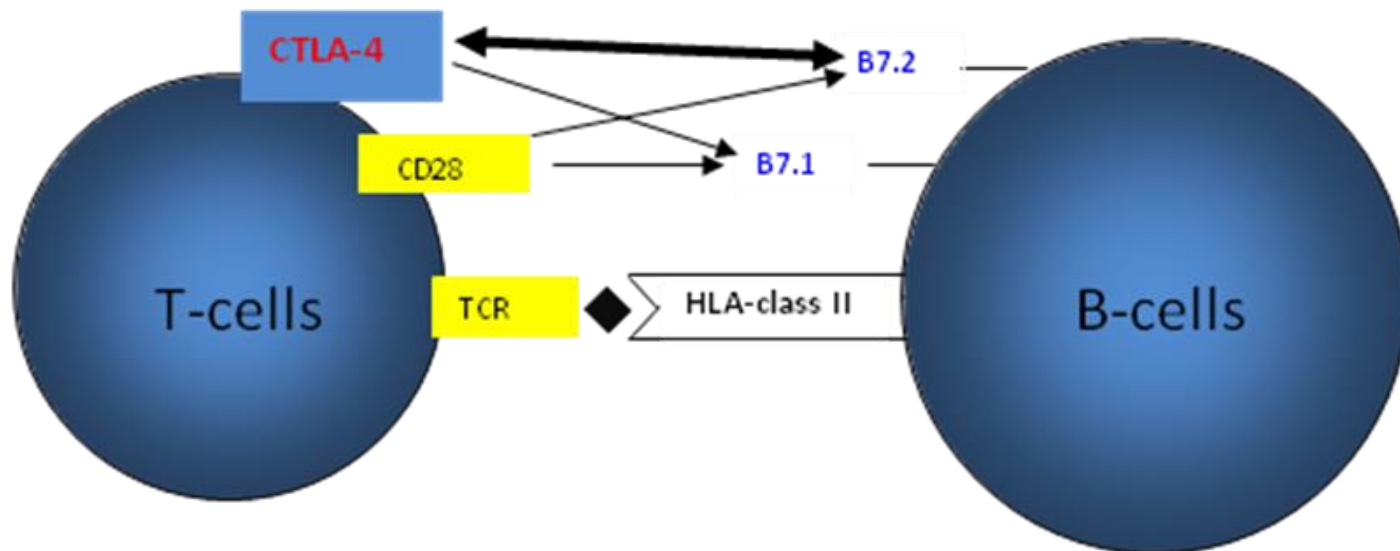


## Effect of interferon beta-1a on B7.1 and B7.2 B-cell expression and its impact on T-cell proliferation

Hui Huang <sup>a,1</sup>, Kouichi Ito <sup>a,2</sup>, Fernando Dangond <sup>b</sup>, Suhayl Dhib-Jalbut <sup>a,\*,2</sup>

<sup>a</sup> Department of Neurology, UMDNJ-Robert Wood Johnson Medical School, 125 Paterson Street, Suite 6200, New Brunswick, NJ 08901, USA

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Contents lists available at ScienceDirect

Journal of Neuroimmunology

journal homepage: [www.elsevier.com/locate/jneuroim](http://www.elsevier.com/locate/jneuroim)



## Comparison of IFN- $\beta$ inducible gene expression in primary-progressive and relapsing-remitting multiple sclerosis

Sridhar Boppana, John E. Mindur, Konstantin E. Balashov, Suhayl Dhib-Jalbut <sup>\*,1</sup>, Kouichi Ito <sup>\*\*1</sup>

*Department of Neurology, Rutgers-Robert Wood Johnson Medical School, Piscataway, NJ 08854, USA*

# PPMS is unresponsive to IFN-B.

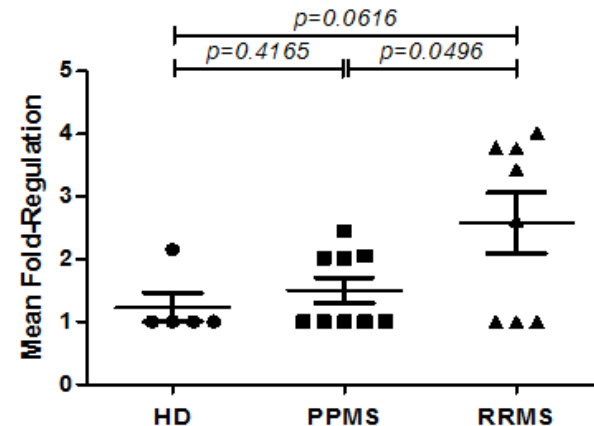
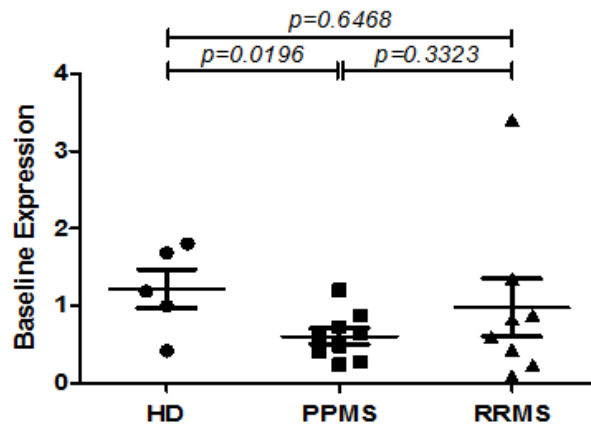
## Why?

# HLA-G is up-regulated in RRMS but not PPMS cells after IFN- $\beta$ treatment *in vitro*

**Table 3.** Induction of IFN- $\beta$  response genes upon *in vitro* treatment with IFN- $\beta$

Gene Symbol	HD (n=5)		PPMS (n=9)		RRMS (n=8)		p-value		
	Fold	SD	Fold	SD	Fold	SD	HD v. PP	PP v. RR	HD v. RR
...									
HLA-C	2.18	± 1.16	1.79	± 0.76	1.40	± 0.78	0.4643	0.3156	0.1748
HLA-F	1.21	± 0.47	1.59	± 0.77	1.66	± 0.96	0.3345	0.8745	0.3527
→ HLA-G	1.23	± 0.52	1.50	± 0.61	2.58	± 1.37	0.4165	<b>0.0496</b>	0.0616
IFI16	3.83	± 1.85	3.60	± 1.69	3.18	± 2.17	0.8169	0.6588	0.5894
IFI27	199.70	± 123.87	50.33	± 53.12	111.57	± 189.09	<b>0.0076</b>	0.3649	0.3781
IFI30	1.62	± 0.87	1.00	± 0.00	1.41	± 1.16	<b>0.0480</b>	0.3037	0.7350
...									

HLA-G



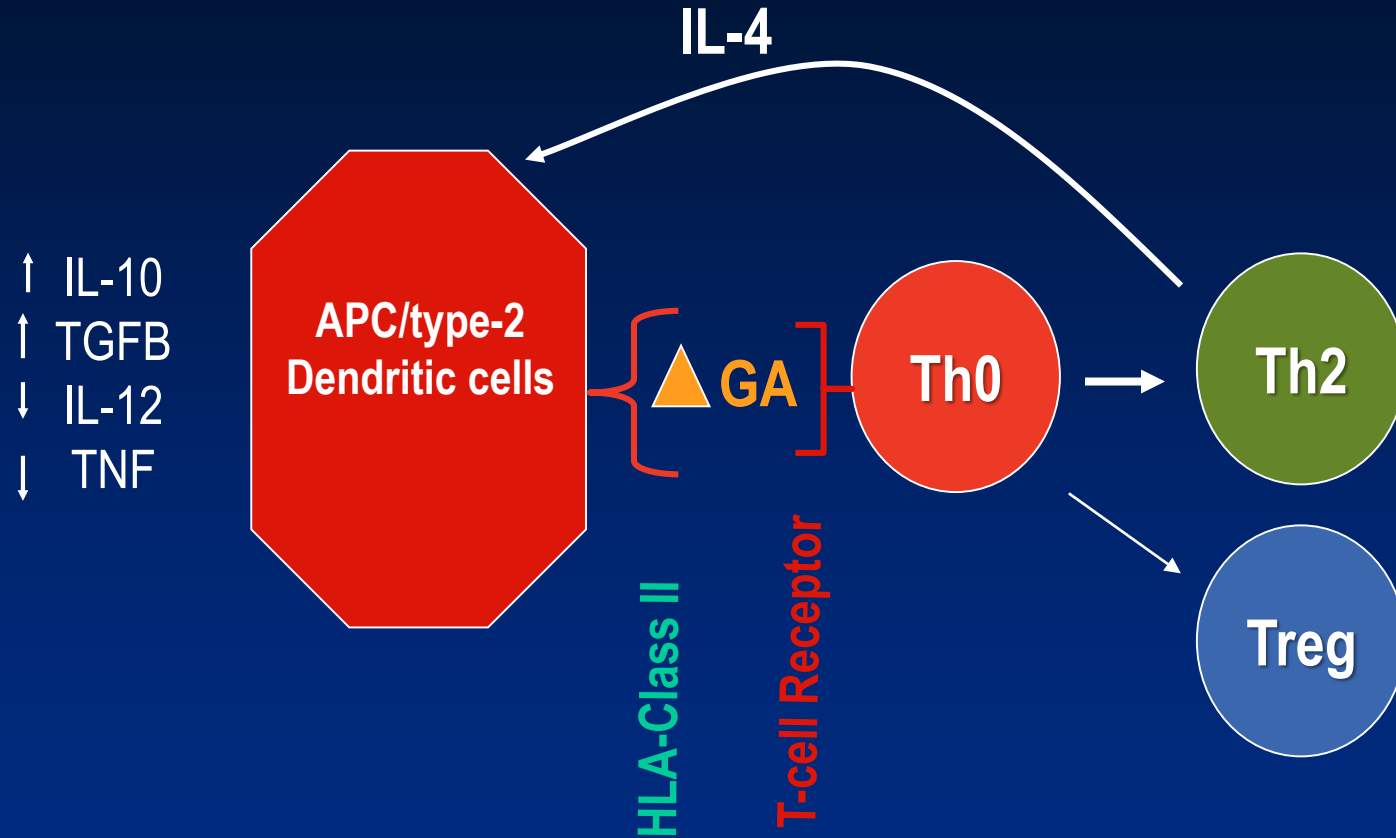
# Potential IFN- $\beta$ Serum Biomarkers

Responders	Non-responders
Increase in IL-10	IL-17F levels > 200pg/ml
Reduction in Th1 cytokines	High baseline IFN- $\beta$ levels
Increased in neurotrophic factors MicroRNA 26a-5p Increased monocytes IFN-I secretion in response to TLR	NAB SNPs (IRF8, IRF5) Increase PSTAT1 and IFNR1 on monocytes at baseline

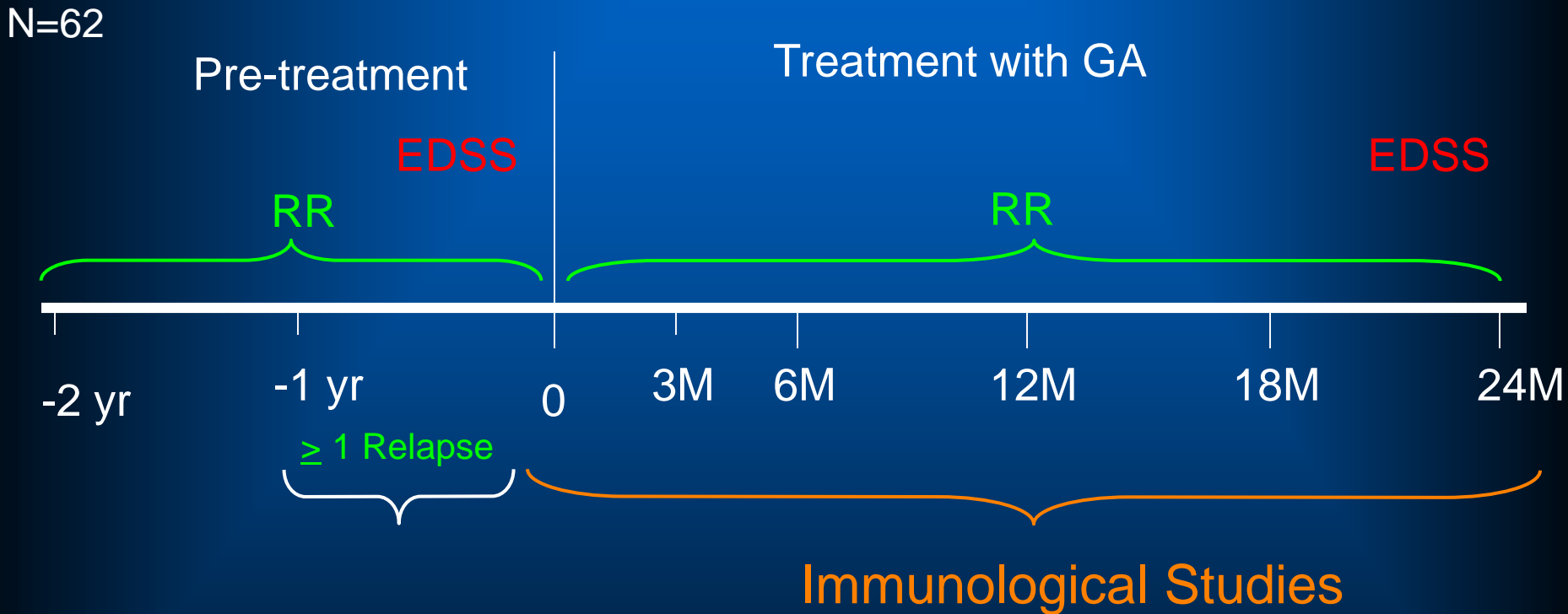
# GLATIRAMER ACETATE



# Glatiramer Acetate Binds to HLA Class II on Antigen Presenting Cells and Induces Type-2 APCs



# Biomarkers of Rx Response to GA: Prospective Study Scheme



# Criteria for Clinical Response

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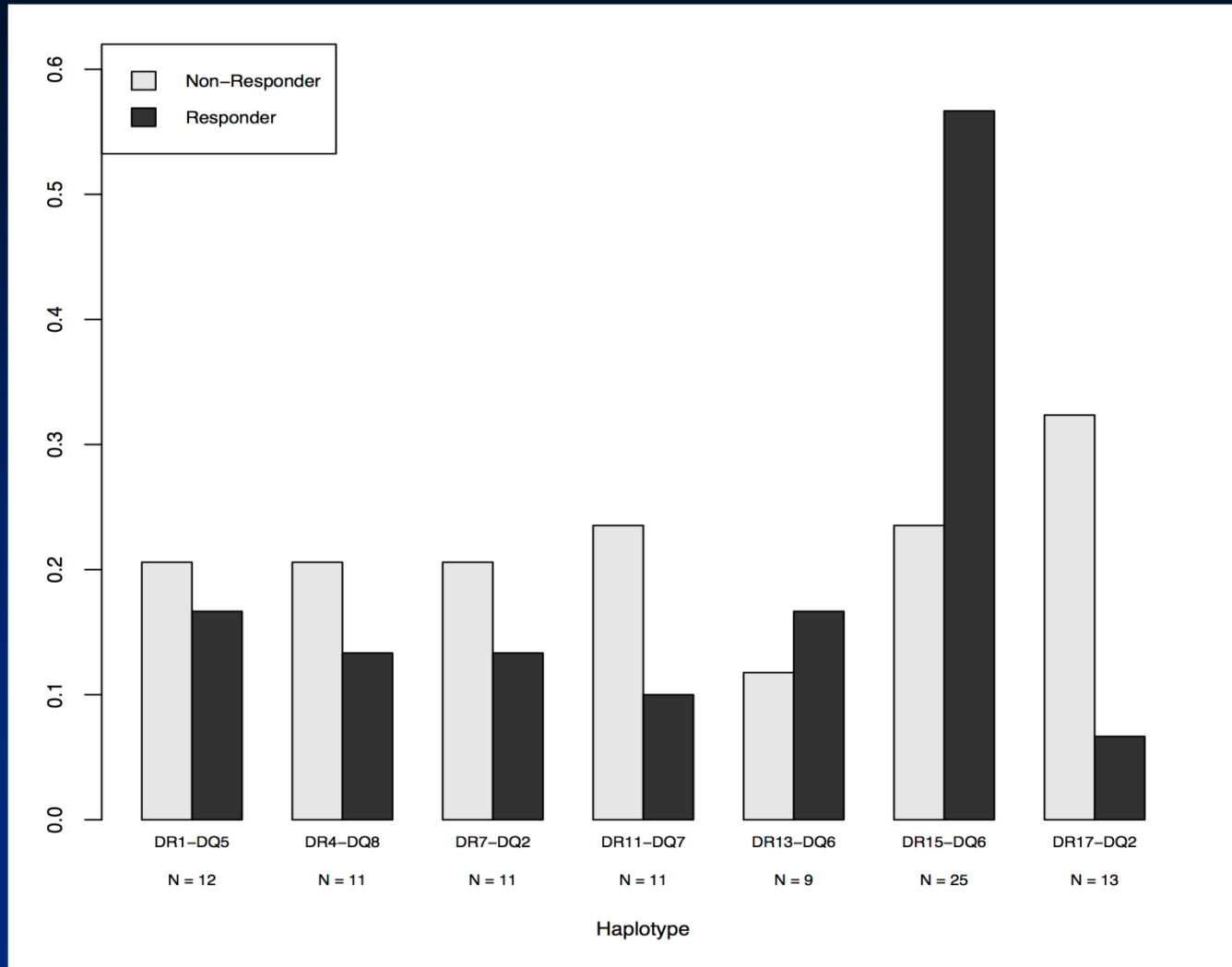
## RESPONDERS

- Patients with no relapses
- With no evidence of disease progression as measured by EDSS

## Non-RESPONDERS

- Patients with  $\geq 1$  Relapse
- With progression in the EDSS of at least 1 point sustained for 6 months

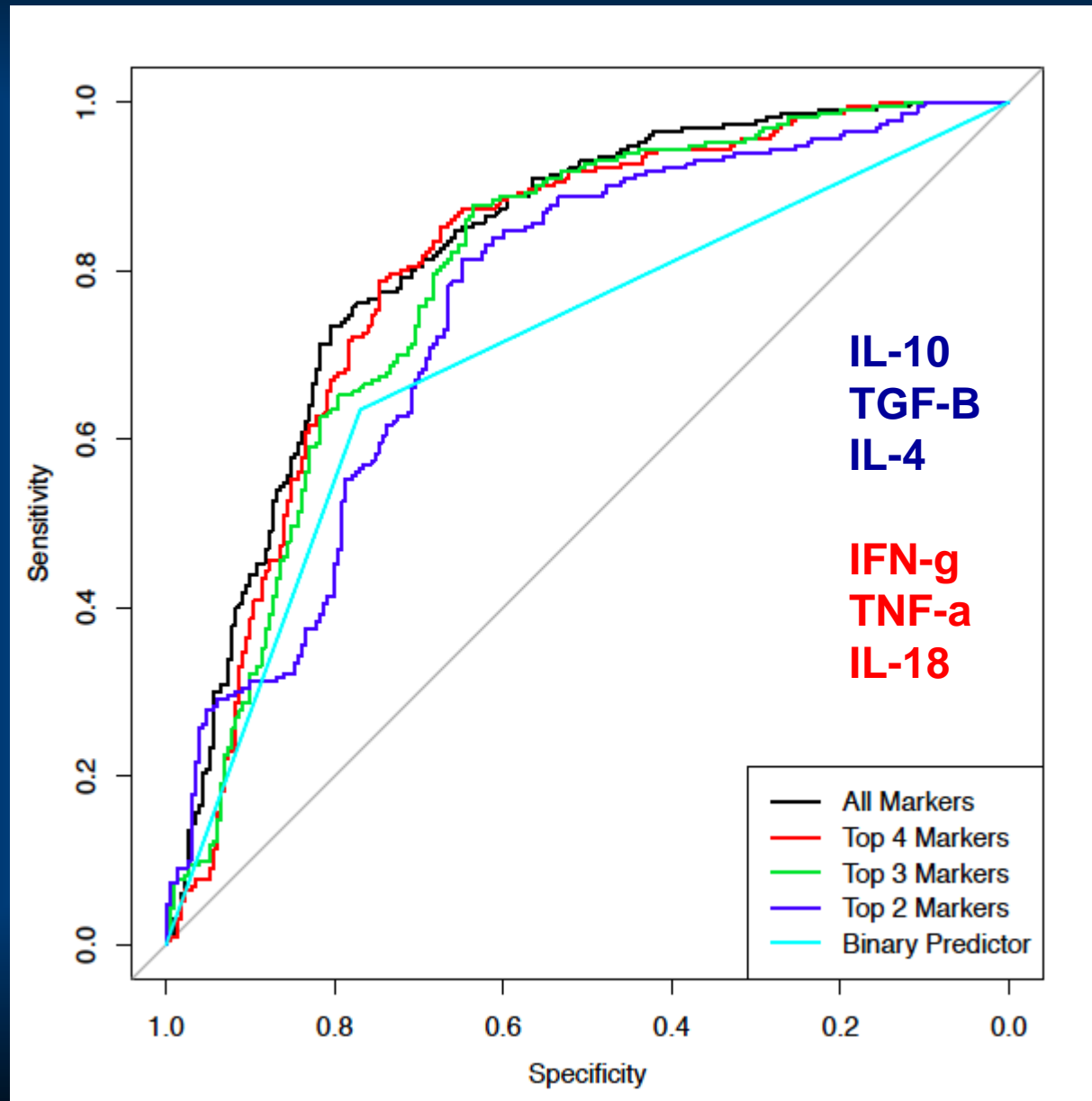
# Response by Haplotype



# DR and DQ Haplotypes Predictors of Clinical Response to GA

PROGNOSTIC PROFILE	HAPLOTYPES	NR / R (%R)
Poor prognostic profile	DR15 - DQ6 absent DR17 - DQ2 present	10 / 2 (16.7%)
Neutral prognostic profile	DR15 – DQ6 present & DR17 – DQ2 present  DR15 – DQ6 absent & DR17 – DQ2 absent	17 / 11 (39.5%)
Good prognostic profile	DR15 – DQ6 present DR17 - DQ2 absent	7 / 17 (70.8%)

# GA induced Cytokine change from baseline; ROC analysis

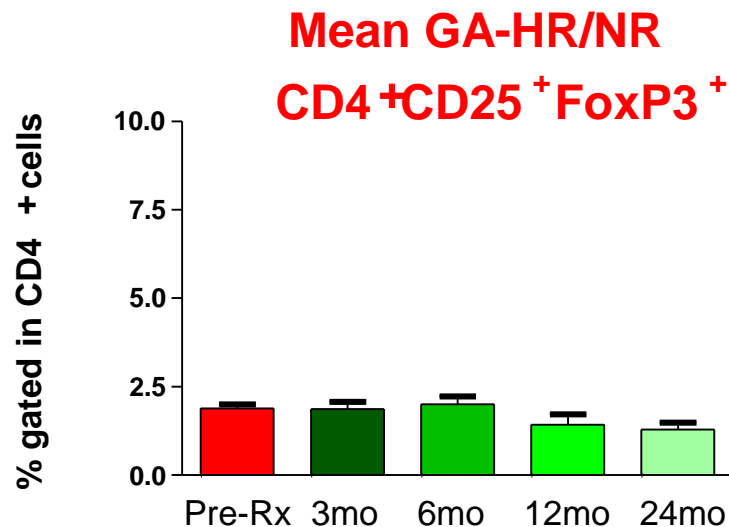
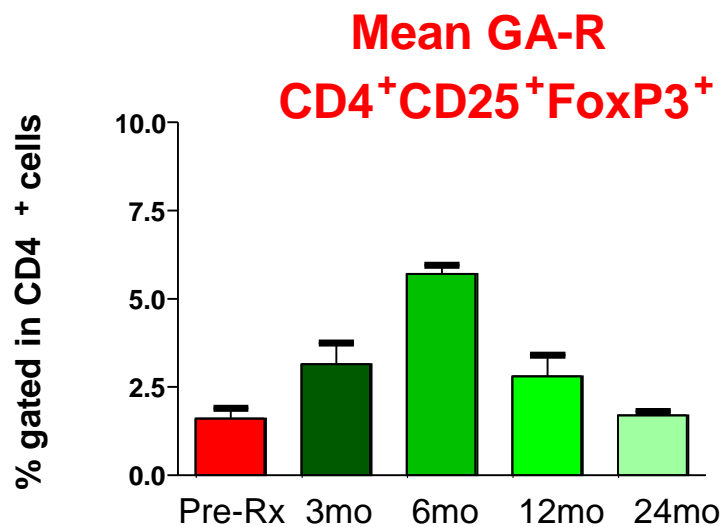


# Binary Cytokine Predictors of response to GA

<b>Clinical outcome</b>	<b><math>\Delta</math>-IL-18&gt;0 <math>\Delta</math>-TGF-B&lt;0</b>	<b><math>\Delta</math>-IL-18&lt;0 <math>\Delta</math>-TGF-B&gt;0</b>
Responders	32.2%	73.4%
Non-Responders	67.8%	26.6%

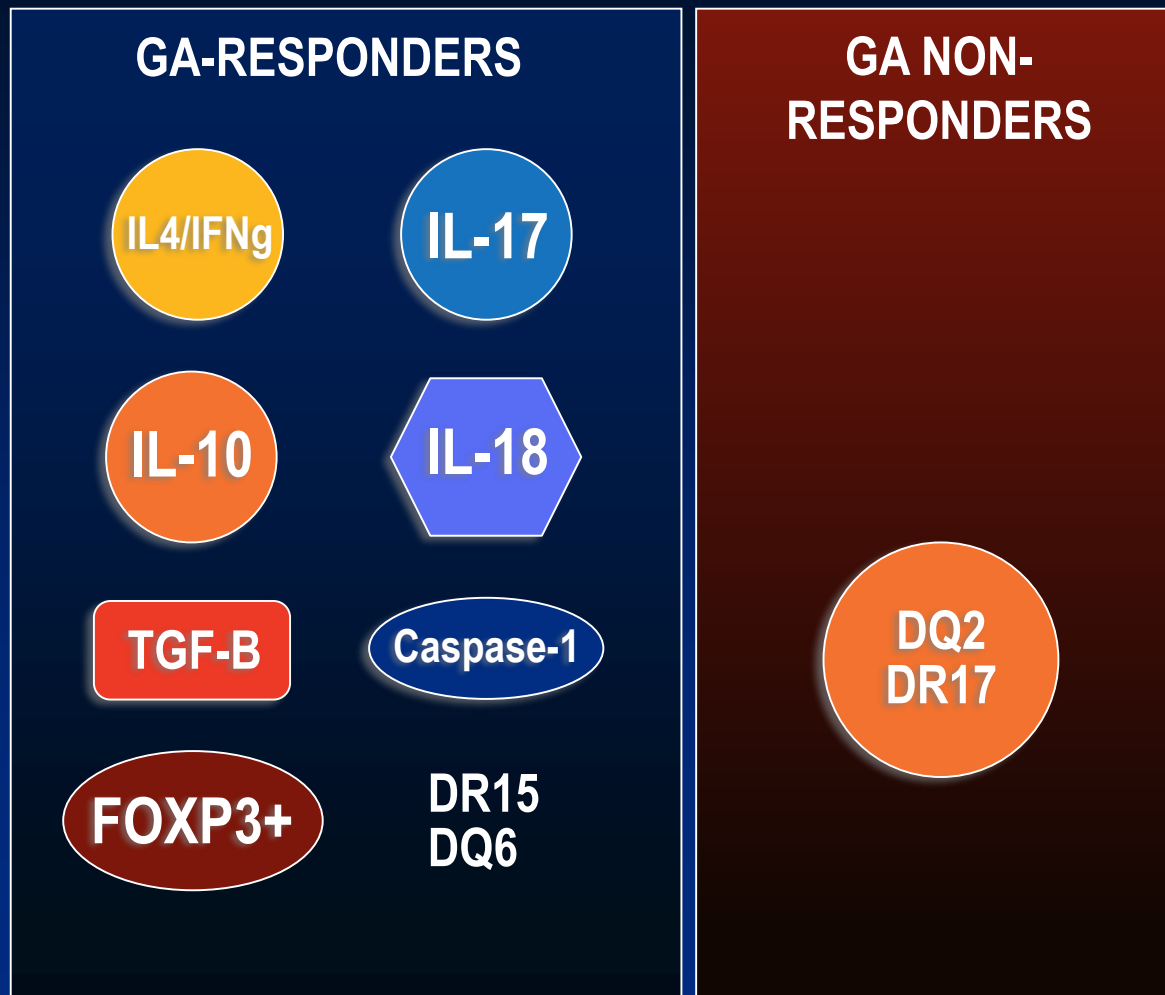
Levels at 6 months compared to base-line  
ROC=0.7

# Induction of CD4<sup>+</sup>CD25<sup>+</sup>FOXP3<sup>+</sup> Tregs During Glatiramer Acetate Treatment





# Summary of Potential GA-Biomarkers

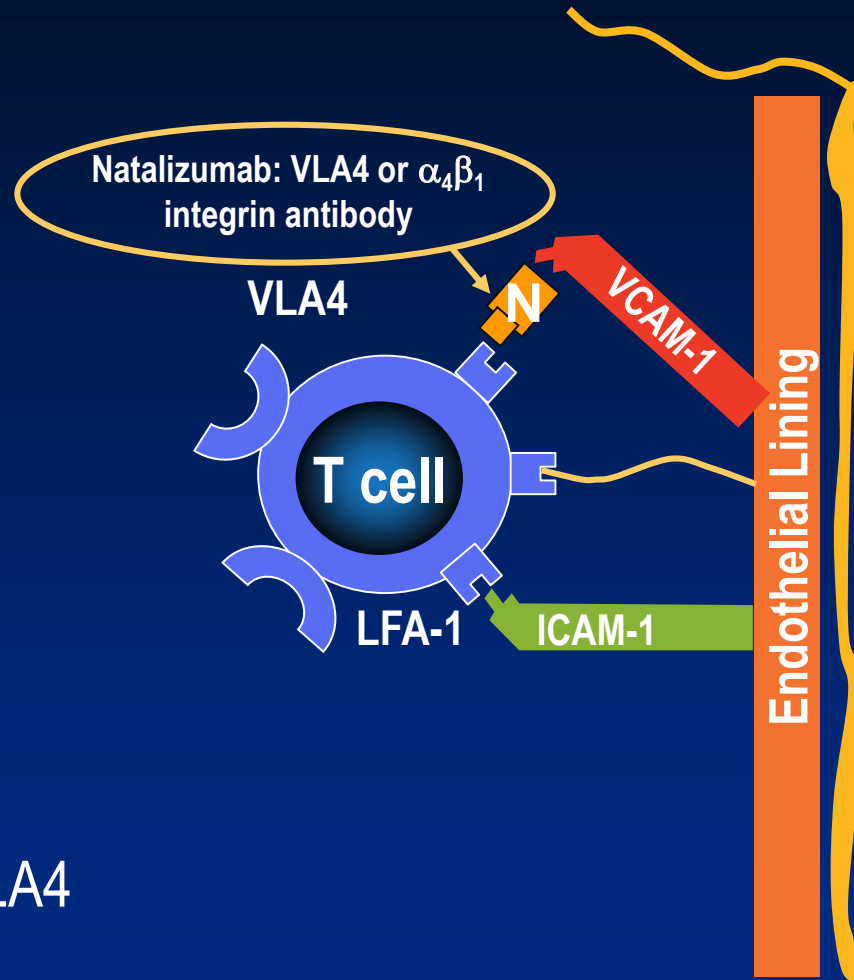


**NATALIZUMAB**

# Natalizumab: Mechanism of Action

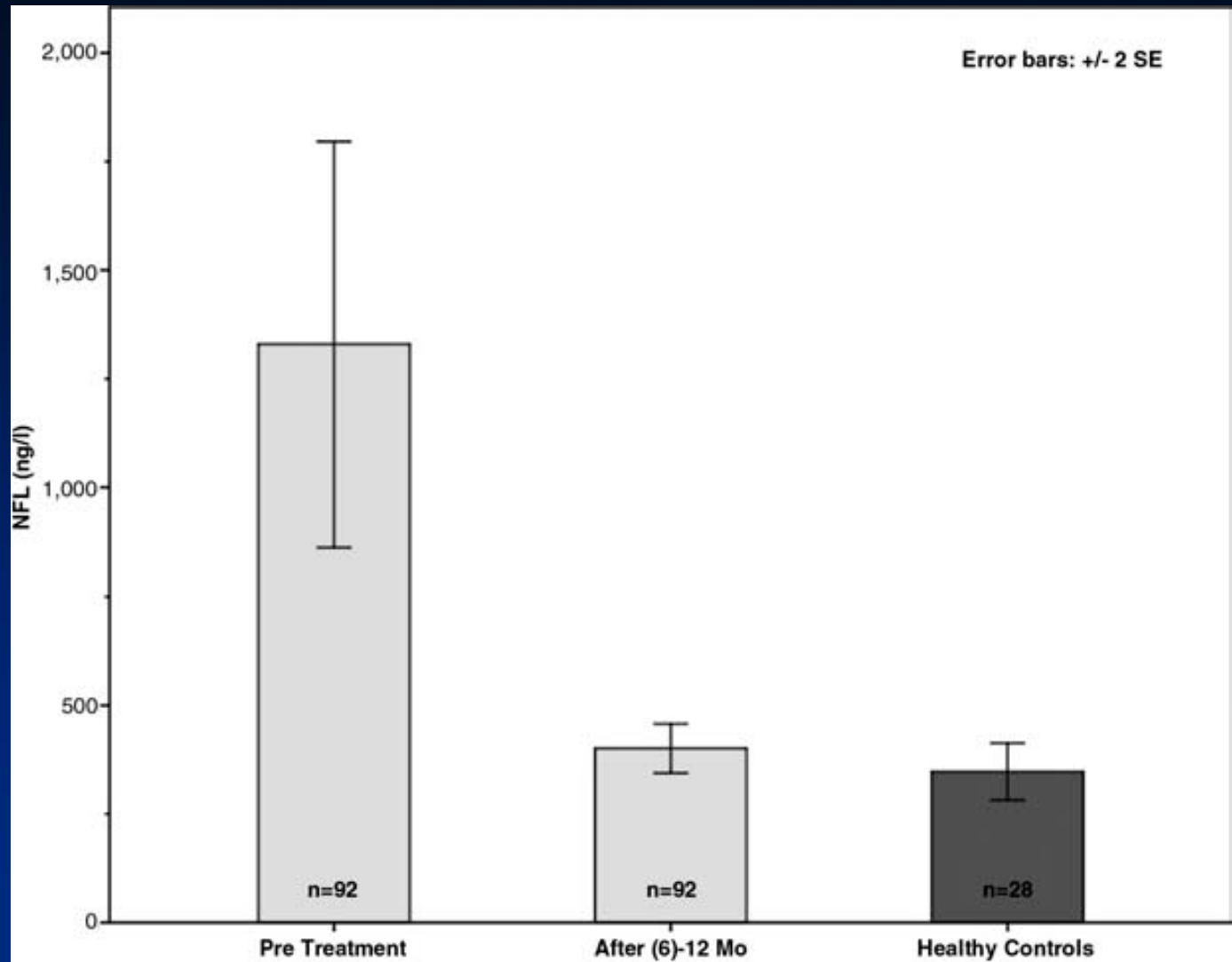
Blood

CNS



$\alpha_4\beta_1$  integrin = VLA4

# Axonal Damage in Relapsing Multiple Sclerosis is Markedly Reduced by Natalizumab



Gunnarsson et al.  
ANN NEUROL  
2011;69:83-89

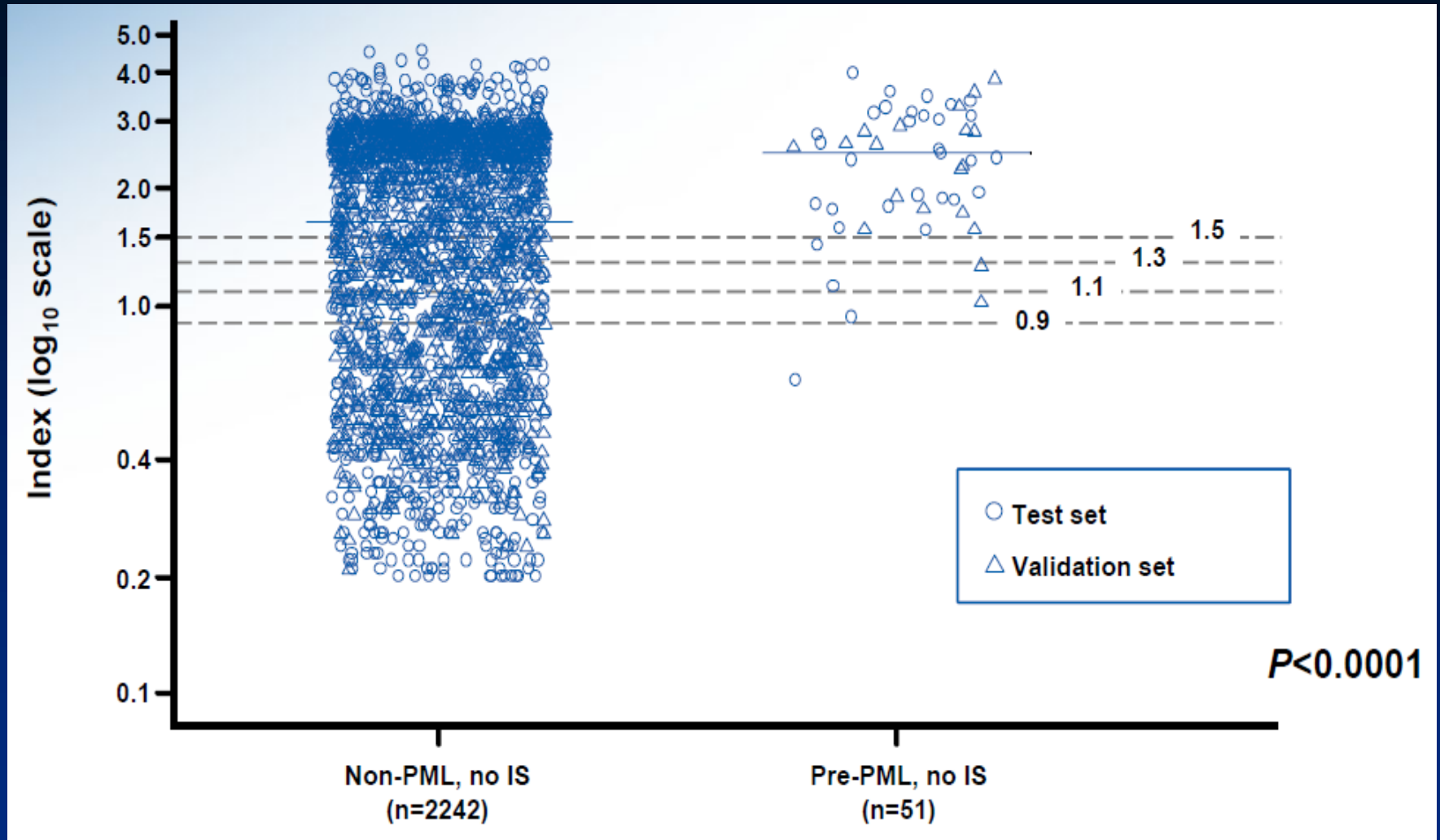
# Potential Exploratory Biomarkers of Newer MS Therapies

Treatment	Tissue	Biomarker
<b>Natalizumab</b>	PB	VLA-4, CD34 cells
	CSF	NFL
<b>Fingolimod</b>	PB	Decreased Naïve and Tcm, Decreased CD4:CD8 ratio, Decreased Th17, Decreased B-cells
	CSF	Decreased T-cells and CD4:CD8 ratio
<b>Rituximab</b>	CSF	Decreased T and B cells
<b>Daclizumab</b>	PB/CSF	Increased NKreg cells
<b>BMT</b>	PB	Decreased TH17

BP: Peripheral Blood; BMT: Bone Marrow Transplant

# **Biomarkers of Treatment Complications**

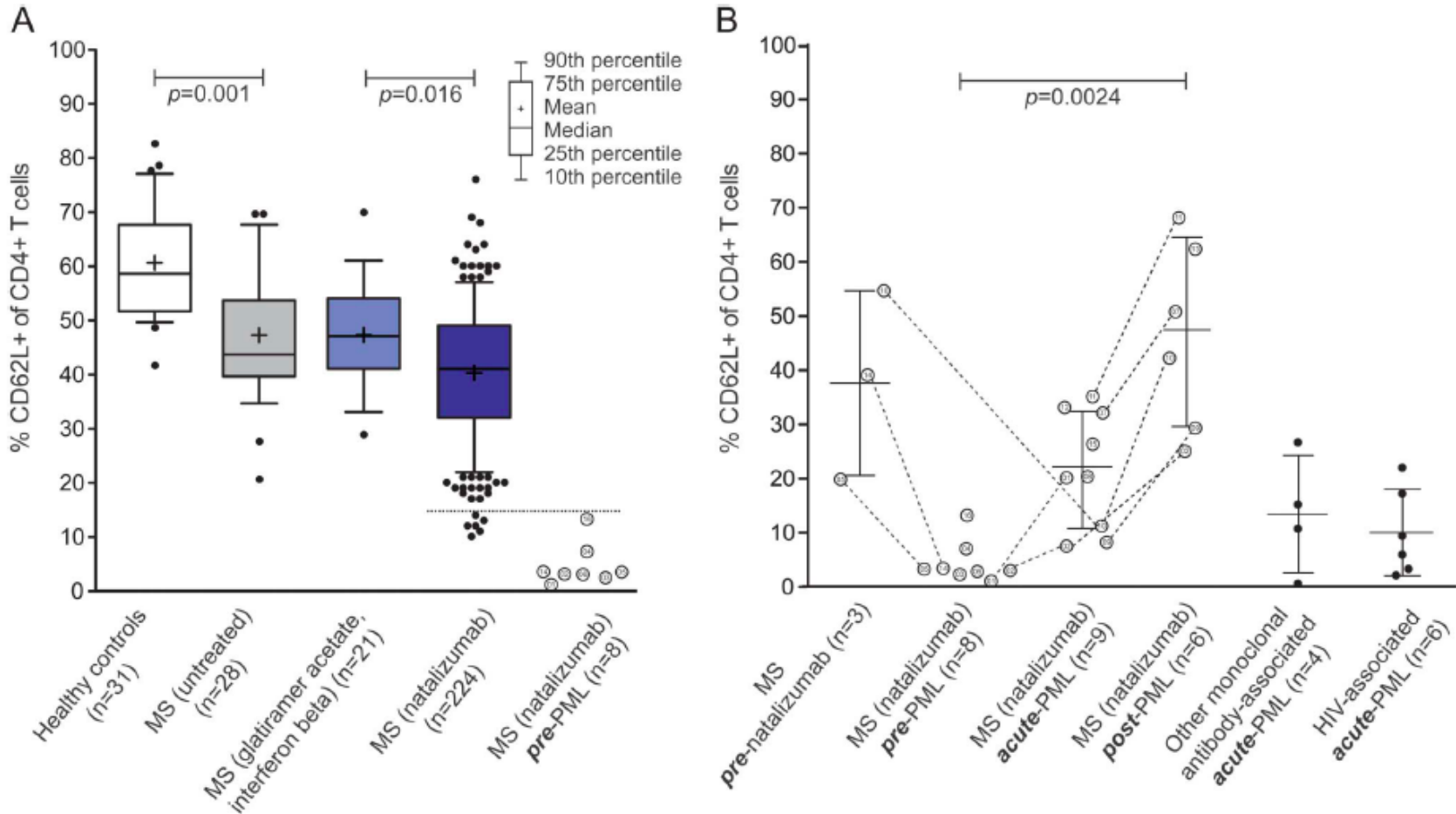
# Anti-JCV Antibody Index Distribution in Anti-JCV Ab+ non-PML and pre-PML Patients with No Prior IS Use



Plavina T, et al. Use of anti-JC virus antibody index to further define risk of PML in anti-JCV antibody-positive TYSABRI-treated patients with MS. Platform presentation presented at 23rd Annual Meeting of European Neurological Society, June 9, 2013; Barcelona, Spain.

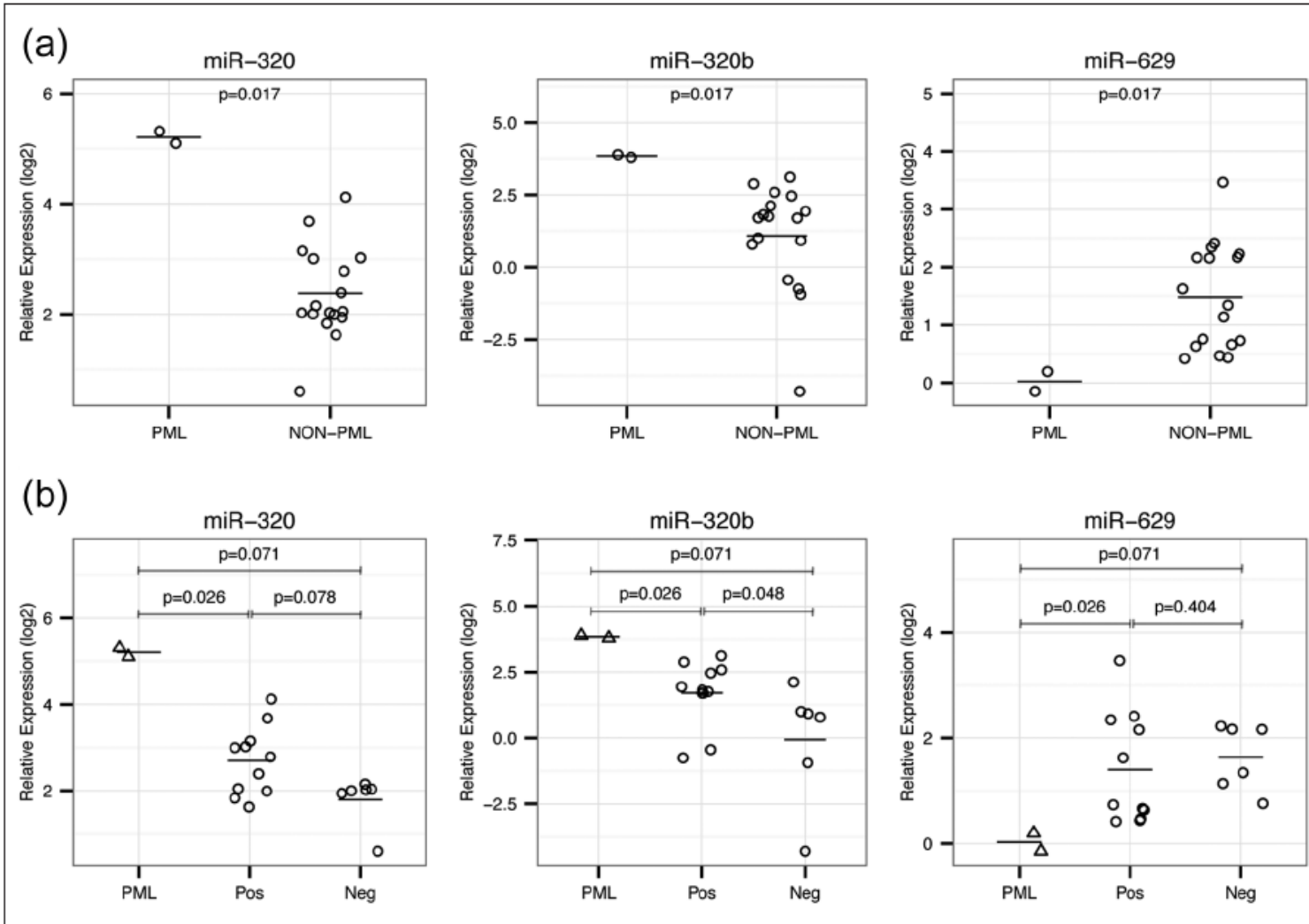
# L-Selectin and Risk of PML in Natalizumab Treated MS Patients

**Figure 1** Surface expression of CD62L and its correlation to progressive multifocal leukoencephalopathy development in multiple sclerosis patients receiving natalizumab therapy

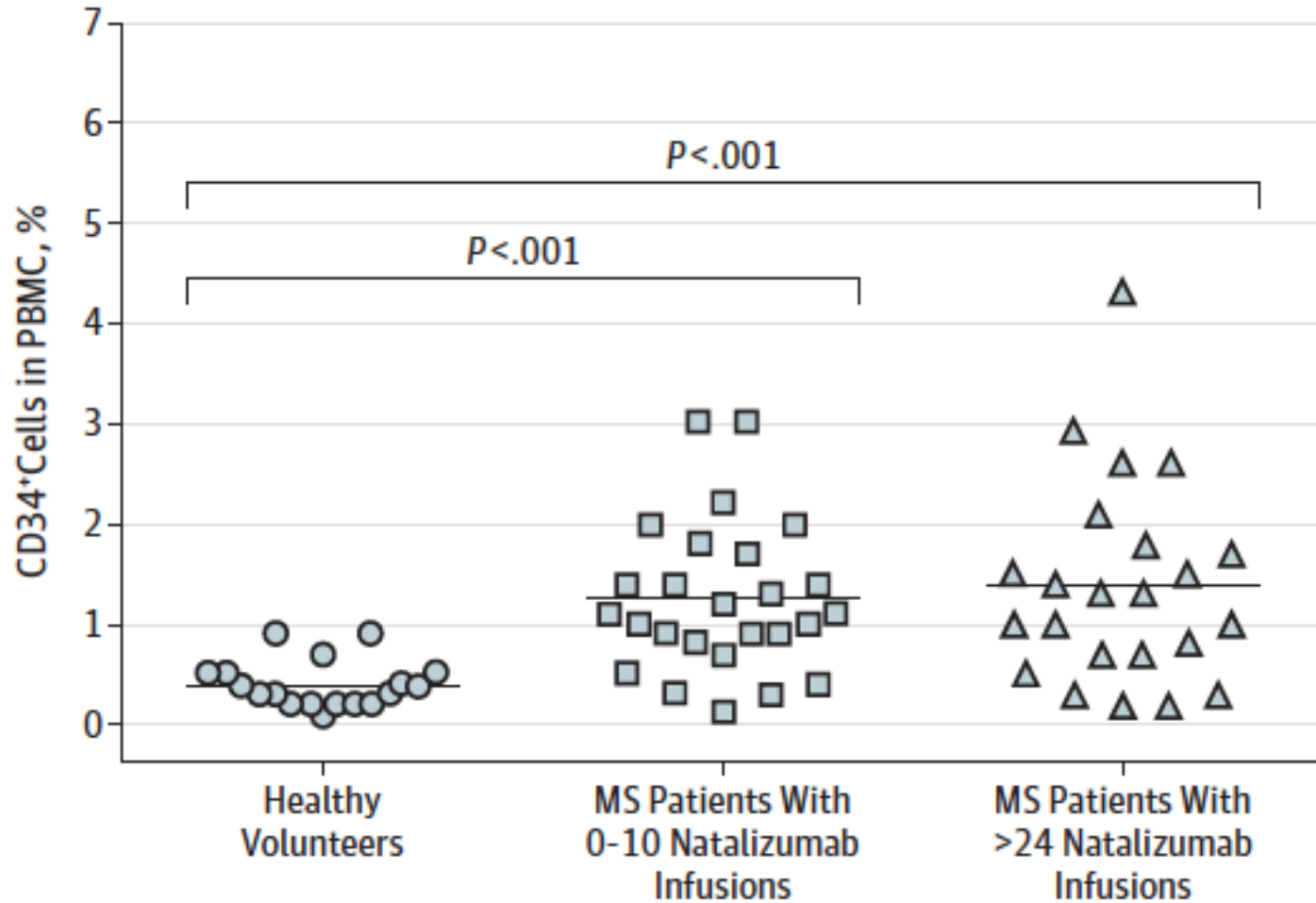




# Blood miRNA expression pattern is a possible risk marker for natalizumab-associated progressive multifocal leukoencephalopathy in multiple sclerosis patients

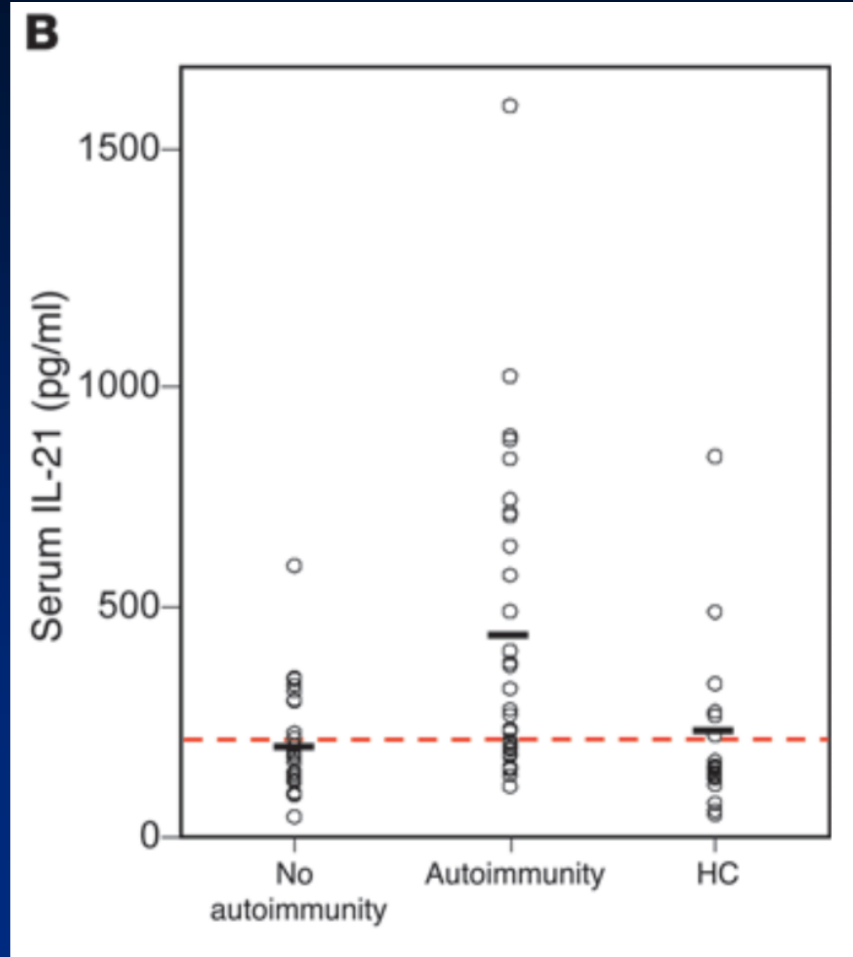
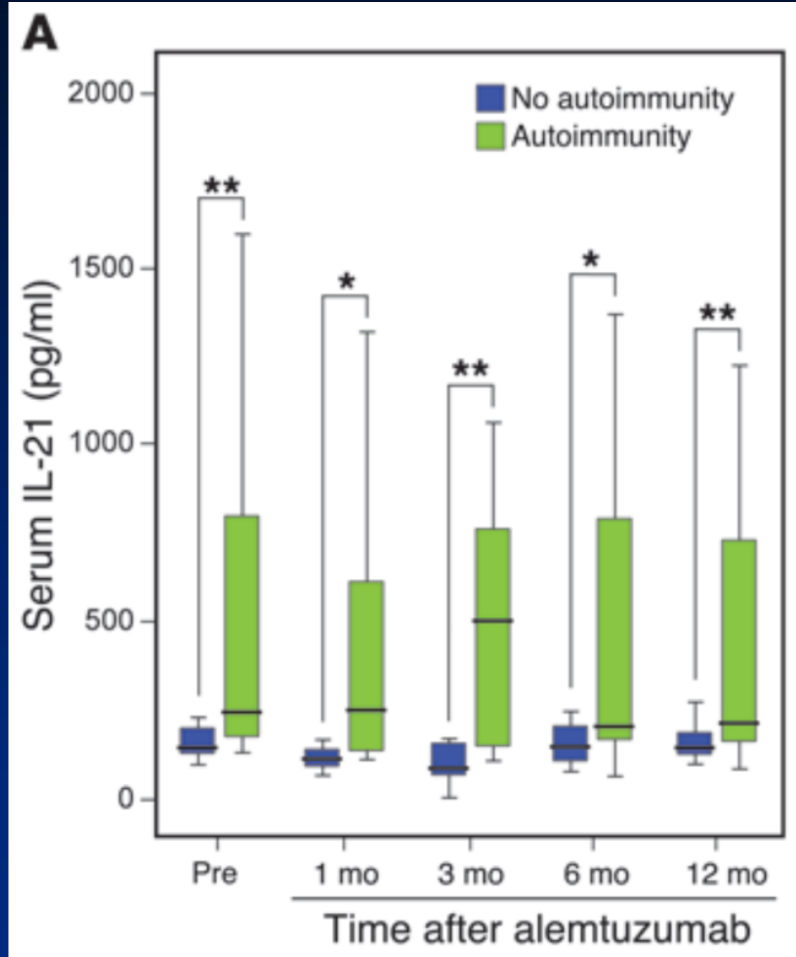


# JC Virus in CD34<sup>+</sup> and CD19<sup>+</sup> Cells in Patients With Multiple Sclerosis Treated With Natalizumab

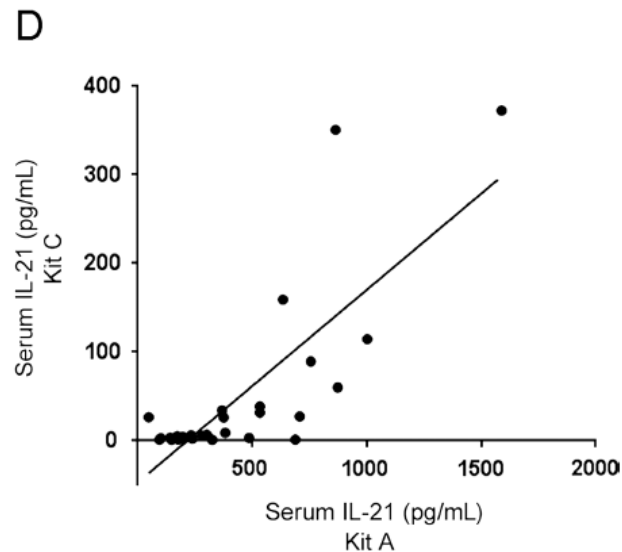
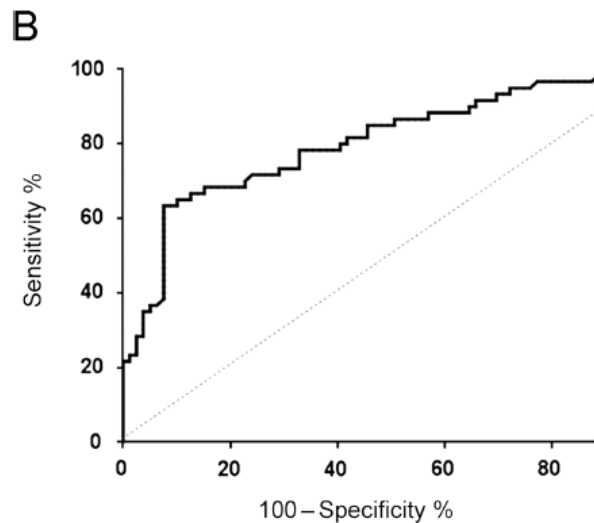
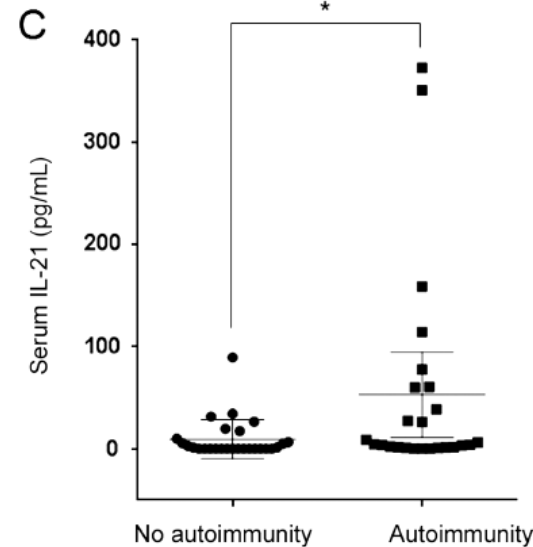
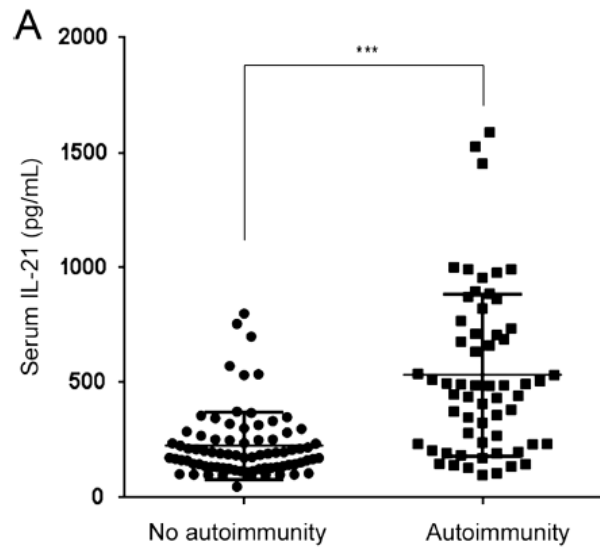


Frohman et al. *JAMA Neurol.* 2014;71(5):596-602.

# Alemtuzumab, serum IL-21 and Autoimmunity



# Alemtuzumab: Predicting Autoimmunity



# Clinically Useful and Potential Biomarkers for Monitoring MS Therapy

Biomarker	Condition
JCV assay	PML risk with Natalizumab
NABs	IFN unresponsiveness
HLA DR & DQ; TGF-B/IL-18	Glatiramer response
IL-10/IL-17/SNIPS	Interferons response
IL-21	Alemtuzumab autoimmunity
CD 56 <sup>Bright</sup>	Daclizumab therapy
MRI	Interferon response

# Acknowledgements

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- Reuben Valenzuela
- Man Chen
- Sumandeeep Sumandeeep
- Jerome Graber
- Steve Buyske
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- NMSS
- DVA
- TEVA Pharmaceuticals
- EMD-SERONO
- BAYER Health
- BIOGEN Idec